

β -CYCLOLAVANDULIC ACID AND SESELIN IN THE ESSENTIAL OIL OF *CARUM ROXBURGHIANUM*

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Key Word Index—*Carum roxburghianum*; Umbelliferae; β -cyclolavandulic acid; seselin.

INTRODUCTION

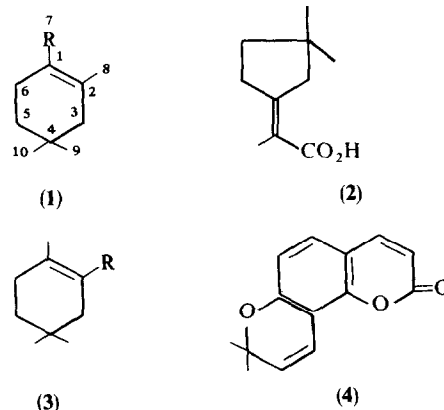
The steam volatile oil of the seeds of *Carum roxburghianum* was reported [1] to contain a keto acid, $C_{10}H_{14}O_3$; mp 105–106° of undetermined structure and another compound, mp 114–115° which was thought to be *p*-isopropyl benzoic acid. We now present evidence for the identification of the compounds as β -cyclolavandulic acid (1; $R = CO_2H$) and seselin (4) respectively.

RESULTS

The essential oil obtained from the seeds of *Carum roxburghianum* was fractionally distilled at 5 mm of Hg and the fractions distilling between 82–104° on standing deposited a crystalline material. This material was dissolved in ether and extracted with dilute sodium bicarbonate solution. The aq. alkaline extract was acidified and extracted with ether. Removal of the solvent gave a solid which on recrystallisation from hexane deposited a white crystalline acid, mp 106–107°. Column chromatography of the oil over Si gel gave the acid in 15–25% yield. The acid analysed for $C_{10}H_{16}O_2$ and its UV absorption at λ_{max}^{EtOH} 223 nm ($\epsilon = 1316$) and IR ($CHCl_3$) ν_{max} 3500–2500, 1680, 1640 cm^{-1} indicated an α,β -unsaturated acid. Its MS showed major peaks at m/e 168 (M^+), 153 ($M^+ - 15$), 140 ($M^+ - 28$), 125 ($M^+ - 43$), 123 ($M^+ - 45$) and 112 ($M^+ - 56$). On catalytic hydrogenation the acid absorbed one mole of hydrogen and gave a dihydro-derivative. m/e 170 (M^+), 137 ($M^+ - 15 - 18$), 110 ($M^+ - 15 - 45$) and 109 ($M^+ - 61$). The PMR ($CDCl_3$, 100 MHz) spectrum revealed the presence of two quarternary methyls at δ 0.92 (6H, s), one methylene at 1.37 (2H, t, $J = 6$ Hz), two allylic methylene at 1.94 (2H, *or*.s) and 2.34 (2H, *m*), one vinylic methyl at 2.06 (3H, *br*.s) and a carboxylic proton at 12.08 (1H, s). The natural abundance ^{13}C -NMR ($CDCl_3$; 25.2 MHz) spectrum in FT mode supported by off resonance decoupled experiments [2, 3] gave nine resonance lines at δ 22.6 (*q*, C-9 and C-10), 23.8 (*t*, C-5), 27.9 (*q*, C-8), 28.7 (*s*, C-4), 34.9 (*t*, C-3), 48.4 (*t*, C-6), 122 (*s*, C-1), 149.5 (*s*, C-2) and 174.3 (*s*, C-7).

The above data pointed to structure 1 ($R = CO_2H$) for the acid which has already been isolated [4, 5] and synthesised [6, 7]. In the absence of a direct comparison, the methyl ester of the acid was ozonised [5] and a keto-ester, PMR ($CDCl_3$; 100 MHz) δ 1.01 (6H, s, Me—C—Me), 2.10 (3H, s, COME), 2.34 (2H, s), 2.81 (2H, *dd*, $J = 8$ Hz), 3.83 (3H, s, CO_2Me) was isolated

which ruled out the possibility of structure 2. In order to distinguish between structure 1 ($R = CO_2H$) and 3 ($R = CO_2H$) for the acid, the diagnostic value of the lanthanide induced shift reagents was employed. Theoretically the induced shift initiated through the functional group at C-1 would give a larger shift of the resonance signal of the C-6 methylene than that of the C-3 methylene in structure 1 and it should be the opposite case in structure 3. The induced paramagnetic shift of the resonance signals of the various protons in the alcohol 1 ($R = CH_2OH$) derived from the acid was observed under varying concentrations of $Eu(fod)_3$ shift reagent. With 41.4 mg of the alcohol in CCl_4 the induced downfield shifts observed for the C-3 and C-6 allylic methylene signals were $\Delta \delta$ 0.12 and 0.34 at 5 mg; 0.23 and 0.74 at 10.8 mg and 0.54 and 1.80 at 25.2 mg concentration of the shift reagent. It was therefore evident that of the two allylic methylenes, the one with a multiplet (C-6), adjacent to another methylene (C-5), had a larger paramagnetic shift compared to the methylene (C-3) having a singlet resonance signal. This confirmed the structure 1 ($R = CO_2H$) for the acid although it might be present as the aldehyde 1 ($R = CHO$) in nature [5] but we could isolate only the acid.



Column chromatography of the essential oil of *Carum roxburghianum* on Si gel with hexane– Et_2O (4:1) gave a fraction which was fluorescent under UV light. The fraction deposited a white crystalline compound, mp 114–115°. The water soluble portion of the essential oil consisted mainly of the same UV fluorescent compound. The total amount of this compound was 13% of the oil. It analysed for $C_{14}H_{12}O_3$; UV λ_{max}^{EtOH} nm: 330, 293,

283, 260, IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1720, 1640, 1600, 1495, 1410, 1300, 1185, 1130, 1025, 855, PMR (CDCl_3 , 100 MHz) δ 1.46 (6H, s), 5.67 (1H, d, $J = 10$ Hz), 6.17 (1H, d, $J = 9$ Hz), 6.66 (1H, d, $J = 8$ Hz), 6.85 (1H, d, $J = 10$ Hz), 7.18 (1H, d, $J = 8$ Hz), 7.57 (1H, d, $J = 9$ Hz), identical with seselin (4). ^{13}C -NMR (CDCl_3 ; 25.2 MHz) gave only eight resonance lines at δ 28.2 (q), 77.3 (d), 112.8 (d), 113.5 (d), 115.3 (d), 127.8 (d), 130 (d), 143.9 (d) which could not be resolved completely. Treatment with sulphuric acid gave umbelliferone [8] which confirmed the structure.

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NEW TERPENE DERIVATIVES FROM *PIQUERIA TRINERVIA**

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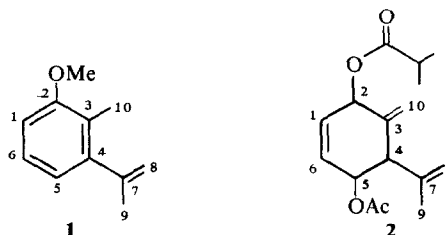
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Key Word Index—*Piqueria trinervia*; Eupatoriaceae; Compositae; new terpene derivatives.

Piqueria trinervia Cav. has been investigated before. Besides the widespread pentayne, santalal and santalol [1] as well as several carquejol derivatives [2] have been reported. A new investigation of the roots yields several new terpenes, all with an unusual carbon skeleton. The ^1H -NMR-data (Tables 1 and 2) lead to the structures 1–4. From the aerial parts, besides 6 also 3 and 4 together with 5 have been isolated. While 1 and 2 are of the same skeleton as carquejol, aldehydes of the isoferulol type like 3–5 up to now only have been isolated from the

Umbelliferae [3]. A compound very similar to 2 has been found in the roots of a *Baccharis* species [4].



* Part 126 in the series 'Naturally Occurring Terpene Derivatives'; Part 125: Bohlmann, F. and Czerson, H. (1978) *Phytochemistry* **17**, 568.

Table 1. ^1H -NMR-data of 1 and 2 (270 MHz, δ -values, CDCl_3)

	1	J (Hz)	2	J (Hz)
1-H	<i>d</i> (br) 6.74	1,6 = 8	<i>ddd</i> 6.02	1,2 = 2.5
2-H	—	—	<i>s</i> (br) 5.78	1,6 = 11
4-H	—	—	<i>d</i> (br) 3.37	1,5 = 2.5
5-H	<i>d</i> (br) 6.83	5,6 = 8	<i>ddd</i> 5.43	4,5 = 5
6-H	<i>dd</i> 7.12	—	<i>d</i> (br) 5.80	5,6 = 3
8-H	<i>dq</i> 5.28	8,8' = 2.3	<i>dq</i> 4.96	5,6 = 3
8'-H	<i>dq</i> 4.79	8,9 = 1.2	<i>s</i> (br) 4.91	—
9-H	<i>dd</i> 1.99	—	<i>s</i> (br) 1.74	—
10-H	<i>s</i> 2.66	—	<i>s</i> (br) 5.30	—
10'-H	<i>s</i> 2.66	—	<i>s</i> (br) 5.21	—
OMe	<i>s</i> 3.80	—	—	—
OAc	—	—	<i>s</i> 2.05	—
OCOR'	—	—	<i>qq</i> 2.57	2', 3' = 7
			<i>d</i> 1.17	—
			<i>d</i> 1.18	—

Table 2. ^1H -NMR-data of 3–5 (270 MHz, δ -values, CDCl_3)

	3	4	5
2-H	<i>d</i> (br) 5.83	<i>d</i> (br) 5.82	<i>d</i> (br) 5.81
3-H	<i>dd</i> 5.62	<i>dd</i> 5.63	<i>dd</i> 5.61
4-H	<i>d</i> (br) 5.76	<i>d</i> (br) 5.74	<i>d</i> (br) 5.74
7-H	<i>s</i> 1.31	<i>s</i> 1.31	<i>s</i> 1.31
8-H	<i>s</i> 1.26	<i>s</i> 1.26	<i>s</i> 1.25
9-H	<i>dd</i> 1.17	<i>dd</i> 2.13	<i>s</i> 2.15
10-H	<i>s</i> 10.25	<i>s</i> 10.25	<i>s</i> 10.24
OCOR'	<i>tq</i> 5.90	<i>tq</i> 6.04	<i>qq</i> 5.72
	<i>s</i> (br) 4.59	<i>s</i> (br) 4.18	<i>d</i> 2,21
	<i>s</i> (br) 2.18	<i>s</i> (br) 2.15	<i>d</i> 1.92
OAc	<i>s</i> 2.13	—	—

J (Hz): 2, 3 = 3, 5; 2, 9 = 1; 3, 4 = 10.5; 9, 10 = 1; 2', 4' = 2', 5' = 1.5.